dase. These facts indicate the virtual absence of acetylmuramic acid residues, unsubstituted by peptide, in these walls. The situation is thus quite different from the hydrolysis of walls of M. lysodeikticus by egg white lysozyme, which treatment alone resulted in liberation of a small percentage of di- and tetrasaccharide (Ghuysen and Salton, 1960; Ghuysen, 1960). The cell wall of S. aureus may therefore be a far more rigid structure than that of M. lysodeikticus. This fact could provide a further explanation for its resistance to egg white lysozyme.

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## Failure of 10 Congeners of myo-Inositol to Support or to Inhibit the Growth of a Cultured Human Cell

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Ten congeners of myo-inositol failed to support the growth of a human cancer cell (KB) even at 10<sup>-4</sup> g/ml, 1000 times the minimal effective concentration of myo-inositol itself. Eight compounds tested at  $10^{-3}$  g/ml failed to inhibit the growth-promoting activity of myo-inositol at  $2 \times 10^{-6}$  g/ml, a ratio of analog to myo-inositol of 500:1.

Interest in the supporting or inhibiting effect of inositol congeners on human cell growth arises from the fact that ordinary (myo) inositol is one of the twentytwo organic compounds (including thirteen amino acids) which are necessary and sufficient for growth of cultured human cells. In the absence of myo-inositol, those defined components, supplemented with dialyzed serum, permit growth only on the addition of serum ultrafiltrate. Experiments in which ninety growth factors were examined showed that myo-inositol was able wholly to replace the ultrafiltrate, while none of the other eighty-nine factors, either separately or together, showed demonstrable activity (Eagle et al., 1956).

Most cultured mammalian cells can synthesize only a fraction of their myo-inositol requirement from glucose (Eagle et al., 1960). One cell line, a mouse fibroblast, not only produced enough for its own survival and growth but released sufficient inositol into the

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medium to permit the parabiotic growth of another and inositol-dependent line. Another cell, a variant of the Hela strain, synthesized marginal amounts, so that exogenous inositol became essential for survival only at inocula of less than 200,000-500,000/ml (Eagle and Piez, 1962). With most cultured mammalian cells, however, exogenous inositol was essential for survival and growth (Eagle et al., 1956), presumably because of the loss of the newly synthesized material to the medium in amounts which exceeded the biosynthetic capacity of the cell (Eagle and Piez, 1962).

A number of recently synthesized inositol analogs and derivatives (McCasland et al., 1954,1961, 1963a,b,c; Shoolery et al., 1961) have now been tested both for their ability to support the growth of an inositolrequiring culture (human carcinoma strain KB) (Eagle, 1955), and for their possible antagonism to myo-inositol itself. The compounds are listed in Table I. None of these substituted for inositol in any concentration up to  $10^{-4}$  g/ml, 100 times the maximally effective concentration of myo-inositol (Eagle et al., 1956), and 1000 times the concentration  $(10^{-7} \text{ g/ml})$  with a partial Further, when eight of growth-promoting action. these compounds were used at 10<sup>-3</sup> g/ml in conjunction

with  $2 \times 10^{-6}$  g/ml of myo-inositol, a 500:1 ratio of analog to inositol, none showed a growth-inhibiting effect (cf. last column in Table I).

TABLE I FAILURE OF TEN CONGENERS OF myo-INOSITOL<sup>a</sup> TO SUPPORT OR TO INHIBIT THE GROWTH OF A CULTURED HUMAN CELL

Common d Montal	Growth <sup>b</sup> in 5 Days when Added at 10 <sup>-4</sup> g/ml to Other- wise Inositol- free Medium	Antagonistic Action of 10 <sup>-3</sup> g/ml to myo- Inositol at 2 × 10 <sup>-3</sup> g/ml (amount of cel- lular growth <sup>b</sup> after 5 days)
Compound Tested	Tree Medium	arter 5 days)
DL-1-Deoxy-epi-inositol	1.30	13.6 ×
(+)-1-Deoxy-allo-inositol		
(2)	0.95	13.8 ×
${(-)\text{-}2\text{-}Deoxy-allo\text{-}inositol}$		······································
23	1.16	_
(-)-3-Bromo-3-deoxy- L-inositol		
$\frac{1}{2}$ Br	1.0	14.4 ×
(-)-3-Chloro-3-deoxy- L-inositol		
2 moshes 2 2 3 Cl	0.89	14.6 ×
(-)-1-Chloro-1-deoxy- neo-inositol		
3 2 CI	0.89	13.5 ×
(-)-1-Iodo-1-deoxy- neo-inositol		
3 2 I	1.27°	13.8 ×

DL(124/5) Stereoisomer of 1,2,4,5-cyclohexane- tetrol		
2	1.14	14.2 ×
meso(13/25) Stereoisomer of 1,2,3,5-cyclohexane- tetrol		
2	0.94	$9.84^{d} \times$
D(126/345) Stereoisomer of 5,6-dimercapto- 1,2,3,4-cyclohexane- tetrol		
SH 3 2 SH	1.0	_
Control with myo- inositol alone	10 <sup>-4</sup> g/ml 12.9 10 <sup>-5</sup> g/ml 12.0 10 <sup>-6</sup> g/ml 11.2	14.0 ×
"The methods of cell of	ultivation the me	dium used and

<sup>a</sup> The methods of cell cultivation, the medium used, and the measurement of cell growth by protein determination have been described in previous communications of Eagle et al. (1956, 1960, 1962). A human cancer cell (KB) culture was used throughout (Eagle, 1955). <sup>b</sup> Referred to inoculum as 1. Insignificant amount of growth, often observed in inositol-free medium.  $^d$  Questionable significance: growth with  $10^{-5}$  g/ml of analog was  $14.3 \times 10^{-5}$  g/ml of analo

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